

# Paratuberculosis: Review of present knowledge

By Michael T. Collins, DVM, PhD

School of Veterinary Medicine University of Wisconsin-Madison, USA

## Introduction

Published literature in the field of paratuberculosis (etiology; *Mycobacterium paratuberculosis*, also known as *Mycobacterium avium* subsp. paratuberculosis) has grown geometrically in the past decade and comprehensive coverage of the topic is beyond the scope of this paper. Readers who desire more in depth information on paratuberculosis should obtain recent reviews and proceedings published on this subject (*Allworth & Kennedy*, *Collins* 2001, *Manning & Collins* 1999 a,b). The website: <http://johnes.org> is another place to obtain current information and references. The goal of this overview is to present some of the major features and recent developments in paratuberculosis.

## Prevalence a slow epidemic

Paratuberculosis occurs in virtually every country of the world. While the infection prevalence is high and best documented in dairy cattle, it has been reported in many different ruminant animals (*Kennedy & Benedictus* 2001). In fact, the range of hosts for this intestinal pathogen appears to be expanding. Scottish investigators found that *M. paratuberculosis* infections can spread from domestic livestock to rabbits and then on to predators of rabbits (*Beard et al.* 2001). The prevalence of paratuberculosis is rising globally and will continue to rise unless control measures are instituted. The coming decades may find paratuberculosis to be the single most economically important single etiology infectious disease of dairy cattle.

## Economic impact

The economic impact of paratuberculosis has been best measured in dairy cattle. In such animals it decreases milk production (*Jones* 1989, *Nordlund et al.* 1996), decreases lifetime production of cows due to premature culling, decreases fertility (*Johnson-Ifeorulundu et al.* 2000), and decreases slaughter value of the carcass (*Whitlock et al.* 1985). An economic impact of potentially greater value is the slowing genetic improvement of herds (or a breeds) due to involuntary culling of genetically valuable animals (*Kennedy & Benedictus* 2001). For herds or flocks, the economic impact of paratuberculosis is not a static parameter, rather it increases as *M. paratuberculosis* infection prevalence increases.

## Pathogenesis

Younger animals are most susceptible to infection by *M. paratuberculosis* infection. This is best documented in dairy cattle (*Gilmour et al.* 1965), but exact measurement of the age-dependent minimal infectious dose has not been reported. Adult cattle can be infected if exposed to large doses of *M. paratuberculosis* (*Rankin* 1961).

The infection starts in the ileum after ingestion of *M. paratuberculosis*. After an incubation period of 1 to 10 years in dairy cattle the clinical manifestations of the disease, most often called Johne's disease, are evident. In cattle this includes diarrhea and weight loss (cachexia). However, in other ruminants diarrhea is less commonly observed and the incubation period

may be shorter. Dairy cattle typically show signs of paratuberculosis when in their second or third lactation. Observation of clinical signs of paratuberculosis at an earlier age indicates a high infection rate in the herd where the animal was born (translating to high infection pressure on young animals).

Infected cows excrete *M. paratuberculosis* in their manure, milk and colostrum. Fecal contamination of feed and drinking water is a very efficient means of infection spread. In the later stages, the infection disseminates beyond the gastrointestinal tract to all internal organs.

### Diagnosis

Clinical signs of paratuberculosis resemble those of many other diseases. At the herd level, signs of paratuberculosis are often simply poor performance or productivity of the herd in spite of good nutrition. Research has provided many new diagnostic tests several of which are available as diagnostic kits helping to provide internationally standardized protocols and reagents for paratuberculosis diagnosis (Collins 1996).

Diagnosis of paratuberculosis by detection of the etiologic agent in clinical samples is favored by many because it is easy to interpret: a positive test confirms a diagnosis. Detection of *M. paratuberculosis* can readily be done by culture or by detection of genetic elements unique to this organism (IS900). Methods of culture have improved and it has been shown that pooling of clinical samples can markedly decrease the cost of culture without significantly compromising sensitivity (Kalis *et al.* 2000). Genetic methods of detection have also improved and techniques to more efficiently extract mycobacterial DNA and/or remove PCR inhibitors help to increase assay sensitivity. Both microbiological and genetic methods of *M. paratuberculosis* detection have the disadvantages of being expensive or slow, or both.

Serological methods of paratuberculosis diag-

nosis have improved. Those based on ELISA technology offers most accuracy per unit cost and can be readily automated for rapid, low cost herd screening. In order to use serological tests for paratuberculosis most effectively, laboratorians and practitioners must learn to deal with diagnostic probabilities rather than certainties and understand the many factors that affect the probability the tested animal has, or does not have an *M. paratuberculosis* infection. Some of the main factors affecting diagnostic probabilities are history of prior cases of paratuberculosis in the herd, clinical status of the animal, and magnitude of the diagnostic test result for those assays that provided quantitative results.

If a herd has never had a case of paratuberculosis, the first step is to confirm a clinical or serological diagnosis of paratuberculosis by culture of *M. paratuberculosis* (from feces or tissues). Once the infection is confirmed in a herd or flock, greater reliance can be placed on positive serological tests. Diagnostic sensitivity of all tests for paratuberculosis is a function of the stage of infection. For example, when applied to animals with clinical signs of Johne's disease the ELISA sensitivity is 85% (Sweeney *et al.* 1995). On *M. paratuberculosis*-infected but clinically normal cattle the ELISA sensitivity is 45% (Socket *et al.* 1992, Sweeney *et al.* 1995). Quantitative interpretation of ELISA results using likelihood ratios is a useful way to guide management of individual animals in a herd and a cost-effective way to selectively cull from the herd those cattle with the highest probability of being infectious.

### Control

Paratuberculosis control programs should be designed to suit the needs and abilities of the business and producer. The more effort expended in changing herd management methods to limit *M. paratuberculosis* infection transmission and the more money spent on diagnostics,

the faster paratuberculosis control can be achieved. Of all control methods, changing herd management, in particular heifer rearing methods, is the single most important factor affecting speed of paratuberculosis control. Test-and-cull programs help accelerate control of paratuberculosis but are not sufficient by themselves to control this infection. Vaccination can decrease the rate of clinical paratuberculosis but it does not prevent *M. paratuberculosis* infection (Kalis *et al.* 2001).

### Prevention / Biosecurity

Herds become infected by buying *M. paratuberculosis*-infected cattle. Pre-purchase testing for Johne's disease is today's standard of veterinary practice. Regardless of method, testing the herd of origin for replacement cattle is much more reliable than testing only the individual purchased cattle. Herd certification programs have started in the U.S. (Ann. 1997), The Netherlands (Benedictus *et al.* 1999), and Australia (Allworth & Kennedy 1999, Kennedy & Allworth 1999). These "certification" programs are formalized efforts to classify herds based on the likelihood of their being *M. paratuberculosis*-infected. Such programs help to limit spread of paratuberculosis by creating a system for cattle buyers purchase animals that have a very low likelihood of having this infection.

### Zoonotic concerns

Crohn's disease and Johne's disease are similar in clinical signs and pathology. New tests for *M. paratuberculosis* find evidence of that this agent infects people with Crohn's disease (Cohavy *et al.* 1999, Collins *et al.* 2000, El Zaatari *et al.* 1999, Hermon-Taylor *et al.* 2000, Naser *et al.* 2000, Olsen *et al.* 2001). Whether the infection is primary (causal) or secondary (incidental or secondary) remains to be determined. Opportunities for human exposure to *M. paratuberculosis* are several. Most attention has

been paid to the idea that dairy products constitute a vehicle for human exposure to *M. paratuberculosis*. Evidence indicates that *M. paratuberculosis* is excreted in raw milk and may resist killing by pasteurization (Stabel *et al.* 2001). Hence, dairy products, in particular those made from dairy products not involving pasteurization in their production, are potential modes of human exposure to *M. paratuberculosis*. However, other modes of exposure deserve equal scrutiny as potential transmission routes. *M. paratuberculosis* is a disseminated infection at the end stages. Contamination of beef products can occur both pre- (via the blood stream) and post- (via fecal contamination) slaughter. As with other microbial contaminants, ground beef have the highest risk of being a vehicle for *M. paratuberculosis* exposure of humans. No published reports quantify the *M. paratuberculosis* numbers in ground beef nor the thermal death rate of *M. paratuberculosis* suspended ground beef. Hence, estimation of the risk of humans' exposure to *M. paratuberculosis* via beef products is nearly impossible.

Water contaminated with manure from domestic animals is another potential route of human exposure that has not been examined. Given that *M. avium* is commonly found in domestic (city) water supplies due to its inherent resistance to chlorine (Falkinham *et al.* 2001, Ann 1997), it is rational to believe that *M. paratuberculosis* too could contaminate surface waters that end up in domestic water supplies and thus expose humans this potential pathogen. A recent cluster of Crohn's disease cases in Mankato, Minnesota, USA was linked to recreational swimming in rivers (Van Kruiningen & Freda 2001).

The debate on the zoonotic potential of *M. paratuberculosis* will continue and likely become more heated in the coming months and years. It is vital that this debate be conducted in an open, public and objective manner. Veteri-

nary medicine must face its dual responsibility to animal agriculture and to the public in controlling this animal health problem and limiting the risk of human exposure to *M. paratuberculosis* through the food chain or environment. When alternative perspectives on the role that *M. paratuberculosis* may play in Crohn's or any other human disease is debated, as at the NKVet Symposium, critical scientific evaluation of study design, methods of analysis, and data interpretation must be applied to keep the debate on the highest plane of scientific objectivity.

## References

- Allworth MB, Kennedy DJ: Progress in national control and assurance programs for ovine Johne's disease in Australia, p. 33-38. In E. J. B. Manning and M. T. Collins (eds.), Proceedings of the Sixth International Colloquium on Paratuberculosis. International Association for Paratuberculosis, Madison, WI, 1999.
- Anonymous: Mycobacterial infections in domestic and wild animals, 350 pages. Office International des Epizooties, Paris, 2001.
- Anonymous: USDA: APHIS:VS, CNAHMS. Johne's disease on U.S. dairy operations, 52 pages. National Animal Health Monitoring System, Ft. Collins, CO, 1997.
- Beard P, Daniels MJ, Henderson DP, Pirie AA, Rudge K, Buxton D, Rhind S, Greig A, Hutchings MR, McKendrick I, Stevenson K, Sharp JM: Paratuberculosis infection in nonruminant wildlife in Scotland. *J. Clin. Microbiol.* 2001, 39, 1517-1521.
- Benedictus G, Verhoeff J, Schukken YH, Hesselink JW: Dutch paratuberculosis programme: History, principles and development, p. 2-15. In E. J. B. Manning and M. T. Collins (eds.), 1999. Proceedings of the Sixth International Colloquium on Paratuberculosis. International Association for Paratuberculosis, Madison, WI.
- Cohavy O, Harth G, Horwitz M, Eggena M, Landers C, Sutton C, Targan SR, Braun J: Identification of a novel mycobacterial histone H1 homologue (HupB) as an antigenic target of pANCA monoclonal antibody and serum immunoglobulin A from patients with Crohn's disease. *Infect. Immun.* 1999, 67, 6510-6517.
- Collins MT: Diagnosis of paratuberculosis. *Veterinary Clinics of North America - Food Animal Practice* 1996, 12, 357-371.
- Collins MT: Task Force on *Mycobacterium paratuberculosis*. *Mycobacterium paratuberculosis*, 62 pages. International Dairy Federation, Brussels, 2001.
- Collins MT, Lisby G, Moser C, Chicks D, Christensen S, Reichelderfer M, Hoiby N, Harms BA, Thomsen OO, Skibsted U, Binder V: Results of multiple diagnostic tests for *Mycobacterium avium* subsp. *paratuberculosis* in patients with inflammatory bowel disease and in controls. *J. Clin. Microbiol.* 2000, 38, 4374-4381.
- El Zaatar FA, Naser SA, Hulten K, Burch P, Graham DY: Characterization of *Mycobacterium paratuberculosis* p36 antigen and its seroreactivities in Crohn's disease. *Curr. Microbiol.* 1999, 39, 115-119.
- Falkinham JO, Norton CD, LeChevallier MW: Factors influencing numbers of *Mycobacterium avium*, *Mycobacterium intracellulare*, and other mycobacteria in drinking water distribution systems. *Appl. Env. Microbiol.* 2001, 67, 1225-1231.
- Gilmour NJL, Nisbet DI, Brotherston JG: Experimental oral infection of calves with *Mycobacterium johnei*. *J. Comp. Path.* 1965, 75:281-286.
- Hermon-Taylor J, Bull TJ, Sheridan JM, Cheng J, Stellakis ML, Sumar N: The causation of Crohn's disease by *Mycobacterium avium* subspecies *paratuberculosis*. *Can. J. Gastroenterol.* 2000, 14, 521-539.
- Johnson-Ifeorulundu YJ, Kaneene JB, Sprecher DJ, Gardiner JC, Lloyd JW: The effect of subclinical *Mycobacterium paratuberculosis* infection on days open in Michigan, USA, dairy cows. *Prev. Vet. Med.* 2000, 46, 171-181.
- Jones RL: Review of the economic impact of Johne's disease in the United States, p. 46-50. In A. R. Milner and P. R. Wood (eds.), Johne's disease. Current trends in research, diagnosis and management. CSIRO, Melbourne, 1989.
- Kalis CHJ, Hesselink JW, Barkema HW, Collins MT: Culture of strategically pooled bovine fecal samples as a method to screen herds for paratuberculosis. *J. Vet. Diagn. Invest.* 2000, 12:547-551.
- Kalis CHJ, Hesselink JW, Barkema HW, Collins MT: Use of long-term vaccination with a killed vaccine to prevent fecal shedding of *Mycobacterium avium* subsp. *paratuberculosis* in dairy herds. *Am. J. Vet. Res.* 2001, 62, 270-274.
- Kennedy DJ, Allworth MB: Progress in national con-

- trol and assurance programs for bovine Johne's disease in Australia, p. 19-26. In E. J. B. Manning and M. T. Collins (eds.), Proceedings of the Sixth International Colloquium on Paratuberculosis. International Association for Paratuberculosis, Madison, WI, 1999.
- Kennedy DJ, Benedictus G: Control of *Mycobacterium avium* subsp. paratuberculosis infection in agricultural species. *Revue Scientifique et Technique* 2001, 20, 151-179.
- Manning EJB, Collins MT: Paratuberculosis in zoo animals, p. 612-616. In M. E. Fowler and R. E. Miller (eds.), *Zoo & Wild Animal Medicine. Current Therapy*. W.B. Saunders, Philadelphia, 1999.
- Manning JB, Collins MT (eds): Proceedings of the Sixth International Colloquium on Paratuberculosis, 720 pages. International Association for Paratuberculosis, Madison, WI, 1999b.
- Naser SA, Schwartz D, Shafran I: Isolation of *Mycobacterium avium* subsp. paratuberculosis from breast milk of Crohn's disease patients. *Am. J. Gastroenterol.* 2000, 95, 1094-1095.
- Nordlund KV, Goodger WJ, Pelletier J, Collins MT: Associations between subclinical paratuberculosis and milk production, milk components, and somatic cell counts in dairy herds. *J. Am. Vet. Med. Assoc.* 1996, 208, 1872-1876.
- Olsen I, Wiker HG, Johnson E, Langeeggen H, Reitan LJ: Elevated antibody responses in patients with Crohn's disease against a 14-kDa secreted protein purified from *Mycobacterium avium* subsp. paratuberculosis. *Scand. J. Immunol.* 2001, 53, 198-203.
- Rankin JD: The experimental infection of cattle with *Mycobacterium johnei* II. Adult cattle inoculated intravenously. *J. Comp. Path.* 1961, 71, 6-9.
- Sockett DC, Conrad TA, Thomas CB, Collins MT: Evaluation of four serological tests for bovine paratuberculosis. *J. Clin. Microbiol.* 1992, 30, 1134-1139.
- Stabel J, Pearce L, Chandler R, Hammer P, Klijn N, Cerf O, Collins MT, Heggum C, Murphy P: Destruction by heat of *Mycobacterium paratuberculosis* in milk and milk products, p. 53-61. In M. T. Collins (ed.), *Mycobacterium paratuberculosis*. International Dairy Federation, Brussels, 2001.
- Sweeney RW, Whitlock RH, Buckley CL, Spencer PA: Evaluation of a commercial enzyme-linked immunosorbent assay for the diagnosis of paratuberculosis in dairy cattle. *J. Vet. Diagn.* 1995, Invest. 7, 488-493.
- Taylor RH, Falkinham JO, Norton CD, LeChevallier MW: Chlorine, chloramine, chlorine dioxide, and ozone susceptibility of *Mycobacterium avium*. *Appl. Env. Microbiol.* 2000, 66, 1702-1755.
29. Van Kruiningen HJ, Freda BJ: A clustering of Crohn's disease in Mankato, Minnesota. *Inflam. Bowel Dis.* 2001, 7, 27-33.
- Whitlock, RH, Hutchison LT, Merkal RS, Glickman LT, Rossiter C, Harmon S, Spencer P, Fetrow J, Bruce J, Benson CE, Dick J: Prevalence and economic consideration of Johne's disease in the northeastern U.S. *Proc. Ann. Mtg. U.S. Anim. Hlth. Assoc.* 1985, 89, 484-490.