Mycobacterium avium subspecies paratuberculosis and Crohn's disease: a systematic review and meta-analysis

Martin Feller, Karin Huwiler, Roger Stephan, Ekkehardt Altpeter, Aijing Shang, Hansjakob Furrer, Gaby E Pfyffer, Thomas Jemmi, Andreas Baumgartner, Matthias Egger

This systematic review assesses the evidence for an association between Mycobacterium avium subspecies paratuberculosis (MAP) and Crohn's disease. We analysed 28 case-control studies comparing MAP in patients with Crohn's disease with individuals free of inflammatory bowel disease (IBD) or patients with ulcerative colitis. Compared with individuals free of IBD, the pooled odds ratio (OR) from studies using PCR in tissue samples was $7 \cdot 01$ (95% CI $3 \cdot 95 - 12 \cdot 4$) and was $1 \cdot 72$ ($1 \cdot 02 - 2 \cdot 90$) in studies using ELISA in serum. ORs were similar for comparisons with ulcerative colitis patients (PCR, $4 \cdot 13$ [$1 \cdot 57 - 10 \cdot 9$]; ELISA, $1 \cdot 88$ [$1 \cdot 26 - 2 \cdot 81$]). The association of MAP with Crohn's disease seems to be specific, but its role in the aetiology of Crohn's disease remains to be defined.

Introduction

Crohn's disease is a chronic inflammatory bowel disease (IBD) of unknown cause, the incidence of which is on the increase in high-income countries.¹ Since the first description of the similarities between Crohn's disease and Johne's disease in cattle in 1913,² it has been argued that *Mycobacterium avium* subspecies *paratuberculosis* (MAP), which causes Johne's disease, might also be a cause of Crohn's disease, and that the dysregulated immune responses are a secondary phenomenon.³-5 Conversely, critics of the mycobacterial theory argue that MAP is a secondary invader rather than a causal factor.6

The association of MAP with Crohn's disease is supported by identification of MAP in patients with Crohn's disease, but not in appropriate controls. The gold standard for detection of MAP is based on isolation of the organism through culture methods. 2,7-10 However, this method is time consuming because of the organism's fastidious nature and slow growth. Molecular and serological methods are widely used alternatives, including immunocytochemistry, 11 nucleic acid hybridisation, 12 and PCR techniques. 13-15 ELISA is commonly used to investigate the immunological evidence of a MAP infection. 16-19

A causal association of MAP with Crohn's disease would have important implications for both prevention and therapy, and is a continuing matter of concern for publichealth agencies. ²⁰⁻²² Since viable MAP organisms are occasionally isolated from commercial pasteurised milk, ²³ the efficacy of some heat-treatment procedures of milk would have to be assessed and improved. Additionally, the search for effective treatment regimens against MAP would need to be intensified.

Our aim was to do a systematic review of case-control studies to assess the evidence that is available on MAP and its association with Crohn's disease.

Methods

Literature search

Literature searches were done in Medline (1966 to December, 2006). Keywords denoting MAP, Crohn's disease or IBD, and the study design were used: "paratuberculosis" (Medical Subject Heading, [MeSH])

or "Mycobacterium paratuberculosis" (MeSH) or "paratuberculosis" (free text); and "Crohn disease" (MeSH) or "inflammatory bowel disease" (MeSH) or "rectal fistula" (MeSH) or "Crohn" (free text); and "case-control studies" (MeSH) or "case-control" (free text). No language restrictions were applied. Additionally, we checked references from relevant publications and review articles.

Eligibility criteria

We included case-control studies if they compared the prevalence of MAP in patients with Crohn's disease with the prevalence in individuals free of IBD by use of PCR or ELISA. Studies comparing patients with Crohn's disease and patients with ulcerative colitis were eligible if they included another comparison group of individuals free of IBD. Studies comparing Crohn's disease patients exclusively with patients with tuberculosis or sarcoidosis were excluded. Studies were excluded if insufficient data were provided to calculate odds ratios (ORs) or if there were two zeros in the 2×2 table. Two reviewers independently assessed eligibility of publications.

Data extraction, outcomes, and definitions

We used a standardised data extraction sheet. Data extraction was done independently by two observers, and any differences were resolved by consensus. We extracted bibliographic, sociodemographic, and clinical data, aspects of study quality, and results. In studies examining several control groups, we chose controls free of IBD for the primary analysis, but, if available, also extracted data for patients with ulcerative colitis. In studies using PCR, the outcome was presence of MAP. In studies using ELISA, the outcome was presence of antibodies against MAP or the antibody titre. If several types of antibodies were assessed, we chose IgG, and if different types of antigens were assessed, we chose the antigen thought to be the most specific for MAP. The specificity of MAP detection tests used was assessed by a specialist in molecular biology (RS), who was provided with only the methods sections of the included publications.

Lancet Infect Dis 2007; 7: 607-13

Institute of Social and Preventive Medicine (ISPM), University of Bern, Bern, Switzerland (M Feller K Huwiler MD, A Shang MD, Prof M Egger MD); Institute for Food Safety and Hygiene. University of Zurich, Zurich, Switzerland (R Stephan DVM); Swiss Federal Office of Public Health, Bern (E Altpeter MD, A Baumgartner PhD); Institute for Infectious Diseases, University of Bern. Bern (H Furrer MD); Department of Medical Microbiology. Cantonal Hospital Lucerne, Lucerne, Switzerland (G E Pfyffer PhD); Swiss Federal Veterinary Office, Berne (T lemmi DVM): and Department of Social Medicine, University of Bristol, Bristol, UK (M Egger)

Correspondence to: Prof Matthias Egger, Institute of Social and Preventive Medicine (ISPM), Finkenhubelweg 11, CH-3012 Bern, Switzerland. Tel +41 31 631 3501; fax +41 31 631 3520; egger@ispm.unibe.ch

Statistical analysis

Study results are presented as ORs with 95% CI. For studies using categorical outcome measures, calculation of an OR was straightforward. If studies measured continuous outcomes, results were converted to ORs by use of the method described by Hasselblad and Hedges.24 This method is based on the fact that, when assuming logistic distributions and equal variances in the two treatment groups, the log OR corresponds with a constant multiplied by the standardised difference between means. An OR above 1.0 indicates a higher prevalence of MAP or higher antibody titres among patients with Crohn's disease compared with controls. The analysis was stratified by the method used (PCR vs ELISA). We calculated the I2 statistic, which describes the percentage of total variation across studies that is caused by heterogeneity rather than chance.25 Low, moderate, and high levels of heterogeneity approximately correspond to I² values of 25%, 50%, and 75%, respectively. There was moderate to high between-study heterogeneity and we

therefore used random-effects meta-analysis to combine the results from different studies. Analyses were done in STATA (version 9.1, STATA Corporation, College Station, TX, USA).

Results

We identified 85 potentially eligible publications, and excluded 29 studies on the basis of title and abstract. 56 studies were examined in detail, of which 28 were excluded for the following reasons: 16 studies reported insufficient information to allow calculation of the OR (including small studies with no positive tests in either group), 19,26-40 two studies were excluded because the serological data from the control group were used to define the threshold for a positive antibody titre (with the consequence that the test was negative by definition in all controls), 41,42 two further studies used ineligible control groups, 43,44 and one study was a duplicate of another study.45 In two cases, publications seemed to report on overlapping sets of patients. The report with the larger

| | Year | Study size | e Country | Control group | | Mean* age (years) | |
|-----------------------------------|------|------------|--------------------|--|-------|-------------------|--|
| | | | | | Cases | Control | |
| Autschbach et al ⁵⁴ | 2005 | 200 | Germany | Intestinal cancer, familiar adenomatous polyposis, other non-IBD-related conditions | 33.9 | 60.8 | |
| Bernstein et al ⁵⁵ | 2003 | 43 | Canada | Healthy | 33.5 | 41.1 | |
| Bull et al ⁵⁷ | 2003 | 71 | UK | Patients undergoing an ileocolonoscopy without clinicopathological diagnosis of Crohn's disease | 32.6 | 55.5 | |
| Clarkston et al ⁵⁹ | 1998 | 32 | USA | Normal colon, IBS, other functional bowel disorder | 37-6 | 45.4 | |
| Dell'Isola et al ¹⁴ | 1994 | 42 | France | Infectious or lymphocytic colitis, polyps, malformations, angiomatosis, rheumatoid purpura, unclassified polyarthritis, autoimmune disease, genetic immunodeficiency, cystic fibrosis | 5-19† | 0.5–171 | |
| Erasmus et al ⁶² | 1995 | 61 | South Africa | Colon cancer | | | |
| Fidler et al ⁶³ | 1994 | 51 | UK | Gut inflammation caused by ileostomies or radiation, carcinomas, vaginoplasties, Meckel's diverticulum | 42 | 45 | |
| Hulten et al ⁷³ | 2001 | 58 | Finland and USA | Diverticular disease, diverticulitis, ischaemic colitis, serosal abscess, cytomegalovirus colitis, acute non-specific colitis, adhesions | | | |
| Lisby et al ⁶⁴ | 1994 | 52 | Denmark | Colonic cancer, peridiverticulitis, ileocaecal lymphoma | 34.5‡ | 67.0‡ | |
| Murray et al ⁶⁵ | 1995 | 24 | New Zealand | Microscopic colitis, ischaemic colitis/diarrhoea, change of bowel habit, colonic polyp, anaemia, diverticular disease, rectal bleeding, IBS | 35.6 | 58.9 | |
| Naser et al ⁶⁶ | 2004 | 43 | USA | Colon cancer, diverticulitis, gastro-oesophageal reflux, healthy individuals | 35.5 | 41.6 | |
| Romero et al ⁶⁷ | 2005 | 18 | USA | Colon cancer | 41.5 | 62-6 | |
| Rowbotham et al ¹⁵ | 1995 | 94 | UK | Rectal bleeding, diarrhoea, changed bowel habit, anaemia, IBS, colonic polyps, ascites, coeliac disease, systemic lupus erythematosus, pseudomembranous colitis | 46.3 | 52.8 | |
| Ryan et al ⁶⁸ | 2002 | 27 | Ireland | Diverticular disease, perianal sinus tissue, colon surgical scar, tubular adenoma of colon, colon carcinoma, colon tuberculosis, mediastinal node sarcoidosis, cholesterol granuloma of breast | | | |
| Sanderson et al ¹³ | 1992 | 80 | UK | Colon cancer, gastric ulcer, sigmoid diverticulitis, small gut sarcoma | 38.1 | 67.7 | |
| Sechi et al ⁴⁶ | 2005 | 71 | Italy | IBS, gastritis, diarrhoea, adenocarcinoma, diverticulitis, screening, colitis, bleeding | | | |
| Suenaga et al ⁶⁹ | 1995 | 26 | Japan | IBS, colon polyps, colon cancer | 39.2 | 54.1 | |
| Tiveljung et al ⁷¹ | 1999 | 22 | Sweden | Colon cancer, colon polyps, functional bowel disorder, ulcerative colitis | 36‡ | 50‡ | |

Table 1: Characteristics of 18 case-control studies that used PCR techniques to detect M avium subspecies paratuberculosis in patients with Crohn's disease and controls

number of patients was included in these instances.^{46,47} One study with a different focus,⁴⁸ two studies that used immunoblotting,^{49,50} and three studies that used culture methods,^{51–53} were also excluded.

We therefore included 28 studies comprising 31 comparisons. 3-18,46,54-74 One study was done in two countries and examined different patient groups, resulting in four comparisons. 60 Reporting and methodological quality of the studies included was variable. Ten of the 28 studies did not report the sex or age distribution of patients and controls, and only 13 studies stated that laboratory staff were blinded to case-control status. 18 of the 31 comparisons used PCR techniques for the detection of MAP in tissue samples and 13 were immunological studies using ELISA.

Among the 18 studies using PCR, the median year of publication was 1999 (range 1992-2005) and the median total sample size was 47 participants (range 18-200; table 1). All studies used categorical outcomes. The median of the mean age of cases was 37.6 years (range 32.6-46.3) and of controls was 54.1 years (range 41.1-67.7). The specimens analysed were tissue samples in all but one study (serum), either obtained by ileocolonoscopy or by surgery. Studies differed regarding the site of biopsies (ileum or colon, inflamed, non-inflamed, or granulomatous tissue) and the depth of tissue extraction (full thickness samples obtained by surgery vs snap samples from ileocolonoscopy). The studies included the following controls: patients with gastrointestinal diseases other than Crohn's disease (predominantly colon diverticulosis, and irritable bowel syndrome) in 17 studies, and healthy individuals in one study. The prevalence of MAP DNA was higher in patients with Crohn's disease than in controls in 16 of 18 studies, resulting in a pooled OR of 7.01 (95% CI 3.95-12.4; figure 1). There was some heterogeneity in study results (I2=44%), which was mainly attributable to the study by Bernstein and colleagues,55 as indicated by a substantial fall in the I² value after exclusion of this study (*I*²=10%). 12 studies compared the prevalence of MAP in Crohn's disease patients with controls free of IBD and with patients with ulcerative colitis. The ORs were similar: 6.88 (3.28-14.4) and 4.13 (1.57-10.9), respectively. We compared the results from the seven studies that used nested PCR with the other studies. Again, ORs were similar: 7.08 (2.19-22.9) and 7.02 $(4 \cdot 12 - 12 \cdot 0)$, respectively.

The median year of publication of studies using ELISA was 1995 (range 1984–2006) and the median sample size was 93 (range 33–685; table 2). The median of the mean age was 36·6 years (range 17·9–44·8) in patients and 37·8 years (range 25·7–42·6) in controls. Outcome measures were continuous in seven studies, ^{16-18,61,70,72,74} and categorical in the remaining studies. All studies analysed serum samples but used different ELISA tests and MAP antigens. The protoplasmic antigen was most frequently used. One study analysed antibodies against a glutathione S-transferase fusion recombinant protein

(part of IS 900). Controls were healthy individuals (seven studies or ten comparisons) or patients with other gastrointestinal conditions (three studies). The prevalence of antibodies against MAP antigens was higher in patients with Crohn's disease than in controls in ten of 13 studies (figure 2). The combined OR was 1.72 (1.02-2.90). All 13 studies also included an ulcerative colitis group; the OR for this comparison was 1.88 (1.26-2.81). Between-study heterogeneity was more pronounced in the comparison

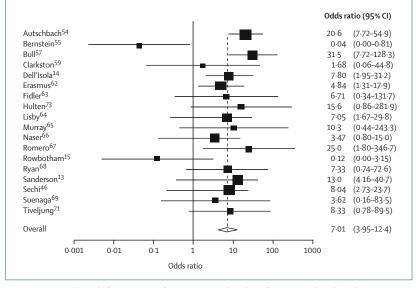


Figure 1: Meta-analysis of 18 comparisons from case-control studies of patients with Crohn's disease versus controls, with PCR in tissue samples or blood to detect M avium subspecies paratuberculosis (MAP)

Odds ratios (ORs) and 95% CI for each study are shown. The size of the square represents the relative weight of each study in the random-effects meta-analysis. The data are displayed on a logarithmic scale. ORs above

1-0 indicate a higher prevalence of MAP in patients with Crohn's disease compared with controls.

| | Year | Study size | Country | Control group | Mean age (years) | |
|---|------|------------|---------|--|------------------|----------|
| | | | | | Cases | Controls |
| Bernstein et al ⁵⁶ | 2004 | 685 | Canada | Healthy | 36.4 | 39.8 |
| Cho et al ⁵⁸ | 1986 | 45 | USA | Duodenal ulcer, diverticulitis or colon cancer | | |
| Collins et al ⁶⁰ | 2000 | 86 | Denmark | Healthy | 39-4 | 42.6 |
| Collins et al ⁶⁰ | 2000 | 111 | Denmark | Healthy | 44.8 | 42.6 |
| Collins et al ⁶⁰ | 2000 | 210 | USA | Healthy | 40-9 | 37.8 |
| Collins et al ⁶⁰ | 2000 | 126 | USA | Healthy | 34.2 | 37.8 |
| Elsaghier et al ⁶¹ | 1992 | 47 | Italy | Healthy | | |
| Nakase et al ⁷⁴ | 2006 | 94 | Japan | Non-inflammatory bowel disease | 35 | 40 |
| Stainsby et al ¹⁶ | 1993 | 76 | UK | Healthy | | |
| Suenaga et al ⁷⁰ | 1999 | 33 | Japan | Healthy | 36.6 | 32.9 |
| Tanaka et al ¹⁸ | 1991 | 93 | UK | Non-inflammatory bowel disease | 17-9 | 25.7 |
| Thayer et al ¹⁷ | 1984 | 123 | USA | Healthy | 38 | 28 |
| Walmsley et al ⁷² =not reported. | 1996 | 61 | UK | Healthy | | |

Table 2: Characteristics of ten case-control studies (13 comparisons) using ELISA tests to detect seropositivity for Mavium subspecies paratuberculosis in patients with Crohn's disease and controls

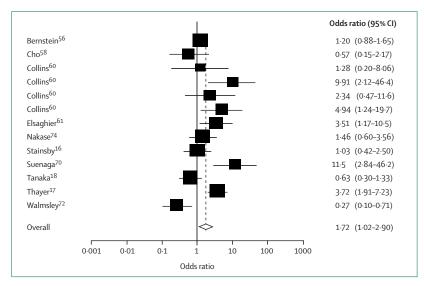


Figure 2: Meta-analysis of 13 comparisons from case-control studies of patients with Crohn's disease versus controls, with ELISA in serum to detect Mavium subspecies paratuberculosis (MAP)

Odds ratios (ORs) and 95% CI for each study are shown. The size of the square represents the relative weight of each study in the random-effects meta-analysis. The data are displayed on a logarithmic scale. ORs above 1-0 indicate a higher prevalence of MAP in patients with Crohn's disease compared with controls.

with controls free of IBD than the comparison with ulcerative colitis patients (*I*²=75% and 44%, respectively).

Discussion

On the basis of 28 case-control studies, this systematic review and meta-analysis shows that tests positive for MAP are substantially more common in patients with Crohn's disease, independently of whether PCR in tissue samples or ELISA in serum is used, or whether patients with Crohn's disease are compared with individuals without IBD or patients with ulcerative colitis.

The results of some of the case-control studies included in our review may have been affected by confounding and bias. In well-designed case-control studies, the source population from which controls are sampled should be the same as that from which cases are also sampled.75 This was not necessarily the case for the studies reviewed here: cases were generally recruited from specialised clinics and controls were either healthy volunteers or patients admitted with other conditions, for example bowel cancer. Furthermore, in case-control studies, it is not possible to establish whether the exposure was present before the onset of the disease or whether it represents an epiphenomenon, which may or may not influence the course of disease once Crohn's disease becomes established. The higher prevalence of positive test results in patients with Crohn's disease could be caused by a higher propensity of inflamed tissue to become infected with MAP.3 We addressed this issue in additional meta-analyses of comparisons with patients with ulcerative colitis. If infection of inflamed mucosa is more likely, one would expect no difference in the prevalence of positive tests, or a smaller difference, when comparing patients with Crohn's disease with patients with ulcerative colitis. Interestingly, we found that tests were more often positive in patients with Crohn's disease, independently of the type of control group. This result supports a specific role of MAP or MAP-like mycobacteria in Crohn's disease, but not in ulcerative colitis. However, it is not proof of a causal role of MAP.

Most PCR studies targeted the MAP DNA insertion element IS900. IS 900 has long been thought to be specific for MAP, but IS 900 elements were also recently found in environmental mycobacteria. Even nested PCR systems (p90/p91; AV1/AV2) gave false-positive results.^{76,77} Therefore, no entirely specific IS 900-based MAP detection exists at present, and a positive result in the tests used in the case-control studies we analysed cannot be equated with MAP infection. The same holds true for the ELISA tests: no MAP-specific test validated for human beings is available.^{50,78} Thus, although positive tests were more common among patients with Crohn's disease than among controls, this is not necessarily because of infection with MAP, but may be attributable to other, MAP-like, bacteria.

There were moderate to high levels of between-study heterogeneity. For case-control studies using PCR, heterogeneity could be attributed to a single outlying study, which contributed little weight to the overall analysis, whereas for the studies using ELISA, the sources of heterogeneity remained unclear. Confounding, bias, and differences in study populations are likely to have contributed to heterogeneity. Our focus was on the association between Crohn's disease and MAP and its specificity, rather than on exploring sources of between-study heterogeneity.

Finally, we excluded 13 studies because neither in patients with Crohn's disease nor in controls was MAP detected. One reason for this could be the method of DNA extraction: in 13 studies without detection of MAP by PCR, only three (23%) used an enzymatic step in combination with a mechanical step or sonication for DNA extraction.

In addition to the mycobacterial theory, other hypotheses exist. For example, Traunmüller79 postulated that environmental or pathogenic mycobacteria (which are able to pass the digestive system because of their acid-fast cell wall) repeatedly stimulate the CD1 system of the intestinal epithelia and lymphoid tissues. Once the gut is pre-immunised, food additives and contaminants, with structures similar to that of mycobacterial lipid antigens, mimic these antigens and boost the immunological reaction.79 If confirmed, this mechanism could lead to an emphasis on dietary treatment. More recently, Marks and colleagues⁸⁰ published data supporting the hypothesis that impaired innate immunity in patients with Crohn's disease predisposes to accumulation of intestinal contents, which can breach the mucosal barrier of the bowel wall. In the absence of adequate numbers of functional neutrophils for the effective clearance of bacteria, bacteria will be taken up by macrophages to form the granulomata and chronic inflammation typical of Crohn's disease. Unfortunately, the potential role of MAP was not examined in these studies.⁸⁰

The association of MAP and Crohn's disease, based on PCR or ELISA testing, is well established and we doubt that important further insights may be gained from additional case-control studies, such as those included in our meta-analysis. Because of the retrospective nature of case-control studies, it is not possible to ascertain whether MAP was present before the onset of the disease or whether it became established secondarily in inflamed tissues. Longitudinal studies are required to answer this question. Because of the low incidence of Crohn's disease, case-control studies nested within cohort studies are the most promising approach to the clarification of the temporal sequence of MAP and Crohn's disease, by use of ELISA in stored serum or, if available, PCR or culture in stored tissue samples. Ideally, such studies should consider genetic factors, in particular mutations of the NOD2 gene, which have been consistently associated with Crohn's disease.1

Another approach to clarifying the role of MAP is the conduct of clinical trials of drug regimens efficacious against MAP. Combination regimens that include macrolide antibiotics and are given for 2 years have been proposed, based on uncontrolled studies that showed substantial benefits. A randomised trial has recently been done in Australia, and data presented at a conference of the Gastroenterology Society of Queensland in June, 2005, indicate that important improvements over placebo were achieved by 16 weeks. Further clinical trials are needed to clarify the place of antimycobacterial combination regimens, including the importance of the duration of therapy and the role of concomitant immunosuppressive therapy.

Conclusions

A causal association of MAP would have important implications for the processing of milk and other dairy products. The occurrence of MAP in milk of productive livestock is well documented, and several studies have shown that viable MAP organisms can survive standard (high-temperature short-time) pasteurisation methods and the processes used for cheese production if high numbers of bacteria are present. ^{23,82–84} There are concerns that use of more thorough heat treatments, which eliminate MAP, would change the organoleptic qualities of milk and adversely affect its taste.

On the basis of this systematic review and metaanalysis, an important, causal role of MAP in the aetiology of Crohn's disease can neither be confirmed nor excluded with certainty. The organism may act as a causative agent, have a role in the context of secondary infection, which may exacerbate the disease, or represent non-pathogenic colonisation. Clearly, the accumulating evidence will need to be updated regularly to allow informed judgments on whether and when public-health action is justified.

Conflicts of interest

We declare that we have no conflicts of interest.

Acknowledgments

This study was funded by the Swiss Federal Office of Public Health and University of Bern.

References

- 1 Shanahan F. Crohn's disease. Lancet 2002; 359: 62-69.
- 2 Dalziel TK. Chronical interstitial enteritis. Br Med J 1913; 2: 1068–70.
- 3 Selby WS. Mycobacterium avium subspecies paratuberculosis bacteraemia in patients with inflammatory bowel disease. Lancet 2004; 364: 1013–14.
- 4 Hermon-Taylor J, Bull T. Crohn's disease caused by Mycobacterium avium subspecies paratuberculosis: a public health tragedy whose resolution is long overdue. J Med Microbiol 2002; 51: 3–6.
- 5 Greenstein RJ. Is Crohn's disease caused by a mycobacterium? Comparisons with leprosy, tuberculosis, and Johne's disease. Lancet Infect Dis 2003; 3: 507–14.
- 6 Chamberlin WM, Naser SA. Integrating theories of the etiology of Crohn's disease. On the etiology of Crohn's disease: questioning the hypotheses. Med Sci Monit 2006: 12: RA27–33.
- 7 Burnham WR, Lennard-Jones JE, Stanford JL, Bird RG. Mycobacteria as a possible cause of inflammatory bowel disease. *Lancet* 1978; 2 (pt 1): 693–96.
- Graham DY, Markesich DC, Yoshimura HH. Mycobacteria and inflammatory bowel disease. Results of culture. Gastroenterology 1987; 92: 436–42.
- Chiodini RJ, Van Kruiningen HJ, Merkal RS, Thayer WR Jr, Coutu JA. Characteristics of an unclassified Mycobacterium species isolated from patients with Crohn's disease. J Clin Microbiol 1984; 20: 066-71
- 10 Chiodini RJ, Van Kruiningen HJ, Thayer WR, Merkal RS, Coutu JA. Possible role of mycobacteria in inflammatory bowel disease. I. An unclassified *Mycobacterium* species isolated from patients with Crohn's disease. *Dig Dis Sci* 1984; 29: 1073–79.
- 11 Kobayashi K, Blaser MJ, Brown WR. Immunohistochemical examination for mycobacteria in intestinal tissues from patients with Crohn's disease. *Gastroenterology* 1989; 96: 1009–15.
- Yoshimura HH, Graham DY, Estes MK, Merkal RS. Investigation of association of mycobacteria with inflammatory bowel disease by nucleic acid hybridization. J Clin Microbiol 1987; 25: 45–51.
- Sanderson JD, Moss MT, Tizard ML, Hermon-Taylor J. *Mycobacterium paratuberculosis* DNA in Crohn's disease tissue. *Gut* 1992; 33: 890–96.
- 14 Dell'Isola B, Poyart C, Goulet O, et al. Detection of Mycobacterium paratuberculosis by polymerase chain reaction in children with Crohn's disease. J Infect Dis 1994; 169: 449–51.
- 15 Rowbotham DS, Mapstone NP, Trejdosiewicz LK, Howdle PD, Quirke P. Mycobacterium paratuberculosis DNA not detected in Crohn's disease tissue by fluorescent polymerase chain reaction. Gut 1995; 37: 660–67.
- Stainsby KJ, Lowes JR, Allan RN, Ibbotson JP. Antibodies to Mycobacterium paratuberculosis and nine species of environmental mycobacteria in Crohn's disease and control subjects. Gut 1993; 34: 371–74.
- 17 Thayer WR Jr, Coutu JA, Chiodini RJ, Van Kruiningen HJ, Merkal RS. Possible role of mycobacteria in inflammatory bowel disease. II. Mycobacterial antibodies in Crohn's disease. *Dig Dis Sci* 1984: 29: 1080–85.
- 18 Tanaka K, Wilks M, Coates PJ, Farthing MJ, Walker-Smith JA, Tabaqchali S. Mycobacterium paratuberculosis and Crohn's disease. Gut 1991; 32: 43–45.
- 19 Kreuzpaintner G, Das PK, Stronkhorst A, Slob AW, Strohmeyer G. Effect of intestinal resection on serum antibodies to the mycobacterial 45/48 kilodalton doublet antigen in Crohn's disease. Gut 1995; 37: 361–66.
- O Food Safety Authority of Ireland. Mycobacterium paratuberculosis. Does it contribute to Crohn's disease? Dublin: Food Safety Authority of Ireland. 2000.

Search strategy and selection criteria

These are described in detail in the Methods section on page 607.

- 21 Advisory Committee on Microbiological Safety of Food. Mycobacterium avium subspecies paratuberculosis and Crohn's disease [ACM493]. London: Advisory Committee on Microbiological Safety of Food, 2001.
- 22 European Union Scientific Committee on Animal Health. Possible links between Crohn's disease and paratuberculosis. Brussels: European Commission, Directorate-General Health and Consumer Protection, 2001.
- 23 Grant IR. Mycobacterium paratuberculosis and milk. Acta Vet Scand 2003; 44: 261–66.
- 24 Hasselblad V, Hedges LV. Meta-analysis of screening and diagnostic tests. Psychol Bull 1995: 117: 167–78.
- 25 Higgins JP, Thompson SG, Deeks JJ, Altman DG. Measuring inconsistency in meta-analyses. BMJ 2003; 327: 557–60.
- 26 Al Shamali M, Khan I, Al Nakib B, Al Hassan F, Mustafa AS. A multiplex polymerase chain reaction assay for the detection of Mycobacterium paratuberculosis DNA in Crohn's disease tissue. Scand J Gastroenterol 1997; 32: 819–23.
- 27 Chiba M, Fukushima T, Horie Y, Iizuka M, Masamune O. No Mycobacterium paratuberculosis detected in intestinal tissue, including Peyer's patches and lymph follicles, of Crohn's disease. J Gastroenterol 1998; 33: 482–87.
- 28 Dumonceau JM, Van Gossum A, Adler M, et al. No Mycobacterium paratuberculosis found in Crohn's disease using polymerase chain reaction. Dig Dis Sci 1996; 41: 421–26.
- 29 Dumonceau JM, Van Gossum A, Adler M, et al. Detection of fastidious mycobacteria in human intestines by the polymerase chain reaction. Eur J Clin Microbiol Infect Dis 1997; 16: 358–63.
- 30 Frank TS, Cook SM. Analysis of paraffin sections of Crohn's disease for Mycobacterium paratuberculosis using polymerase chain reaction. Mod Pathol 1996; 9: 32–35.
- 31 Fujita H, Eishi Y, Ishige I, et al. Quantitative analysis of bacterial DNA from mycobacteria spp, *Bacteroides vulgatus*, and *Escherichia coli* in tissue samples from patients with inflammatory bowel diseases. *J Gastroenterol* 2002; 37: 509–16.
- 32 Gibson J, Riggio M, McCreary C, Lennon A, Toner M. Looking for Mycobacterium paratuberculosis DNA by polymerase chain reaction (PCR) in orofacial granulomatosis (OFG) and oral Crohn's disease tissue in an Irish population. Ir Med J 2000; 93: 218.
- 33 Kallinowski F, Wassmer A, Hofmann MA, et al. Prevalence of enteropathogenic bacteria in surgically treated chronic inflammatory bowel disease. *Hepatogastroenterology* 1998; 45: 1552–58
- 34 Kanazawa K, Haga Y, Funakoshi O, Nakajima H, Munakata A, Yoshida Y. Absence of Mycobacterium paratuberculosis DNA in intestinal tissues from Crohn's disease by nested polymerase chain reaction. J Gastroenterol 1999; 34: 200–06.
- 35 Kreuzpaintner G, Kirschner P, Wallner A, et al. Mycobacteria of Runyon groups I, II and IV do not play an aetiological role in Crohn's disease. Eur J Gastroenterol Hepatol 1995; 7: 1177–82.
- 36 Mishina D, Katsel P, Brown ST, Gilberts EC, Greenstein RJ. On the etiology of Crohn disease. Proc Natl Acad Sci U S A 1996; 93: 9816–20.
- 37 Riggio MP, Gibson J, Lennon A, Wray D, MacDonald DG. Search for Mycobacterium paratuberculosis DNA in orofacial granulomatosis and oral Crohn's disease tissue by polymerase chain reaction. Gut 1997; 41: 646–50.
- 88 Rosenberg W, Bell J, Jewell D. Mycobacterium paratuberculosis DNA cannot be detected in Crohn's disease tissues. Gastroenterology 1991; 100: A611.
- 39 Cellier C, De Beenhouwer H, Berger A, et al. Mycobacterium paratuberculosis and Mycobacterium avium subsp. silvaticum DNA cannot be detected by PCR in Crohn's disease tissue. Gastroenterol Clin Biol 1998: 22: 675–78.
- 40 Sechi LA, Mura M, Tanda E, Lissia A, Fadda G, Zanetti S. Mycobacterium avium sub. paratuberculosis in tissue samples of Crohn's disease patients. New Microbiol 2004; 27: 75–77.
- 41 Brunello F, Pera A, Martini S, et al. Antibodies to Mycobacterium paratuberculosis in patients with Crohn's disease. Dig Dis Sci 1991; 36: 1741–45.
- 42 Cohavy O, Harth G, Horwitz M, et al. Identification of a novel mycobacterial histone H1 homologue (HupB) as an antigenic target of pANCA monoclonal antibody and serum immunoglobulin A from patients with Crohn's disease. *Infect Immun* 1999; 67: 6510–17.

- 43 Ikonomopoulos JA, Gorgoulis VG, Kastrinakis NG, et al. Sensitive differential detection of genetically related mycobacterial pathogens in archival material. Am J Clin Pathol 2000; 114: 940–50.
- 44 Quirke P, Dockey D, Taylor G, et al. Detection of Mycobacterium paratuberculosis in inflammatory bowel disease. Gut 1991; 32: A572.
- 45 Sanderson J, Moss M, Malik Z, Tizard M, Green E, Hermon-Taylor J. Polymerase chain reaction detects Mycobacterium paratuberculosis in Crohn's disease tissue extracts. Gut 1991; 32: A572.
- 46 Sechi LA, Gazouli M, Ikonomopoulos J, et al. Mycobacterium avium subsp paratuberculosis, genetic susceptibility to Crohn's disease, and Sardinians: the way ahead. J Clin Microbiol 2005; 43: 5275–77.
- 47 Sechi LA, Scanu AM, Molicotti P, et al. Detection and isolation of Mycobacterium avium subspecies paratuberculosis from intestinal mucosal biopsies of patients with and without Crohn's disease in Sardinia. Am J Gastroenterol 2005; 100: 1529–36.
- 48 Polymeros D, Bogdanos DP, Day R, Arioli D, Vergani D, Forbes A. Does cross-reactivity between Mycobacterium avium paratuberculosis and human intestinal antigens characterize Crohn's disease? Gastroenterology 2006; 131: 85–96.
- 49 Naser SA, Hulten K, Shafran I, Graham DY, el Zaatari FA. Specific seroreactivity of Crohn's disease patients against p35 and p36 antigens of M avium subsp paratuberculosis. Vet Microbiol 2000; 77: 497–504.
- 50 el Zaatari FA, Naser SA, Hulten K, Burch P, Graham DY. Characterization of Mycobacterium paratuberculosis p36 antigen and its seroreactivities in Crohn's disease. Curr Microbiol 1999; 39: 115–19.
- 51 Moss MT, Sanderson JD, Tizard ML, et al. Polymerase chain reaction detection of Mycobacterium paratuberculosis and Mycobacterium avium subsp silvaticum in long term cultures from Crohn's disease and control tissues. Gut 1992; 33: 1209–13.
- 52 Schwartz D, Shafran I, Romero C, et al. Use of short-term culture for identification of Mycobacterium avium subsp. paratuberculosis in tissue from Crohn's disease patients. Clin Microbiol Infect 2000; 6: 303–07.
- 53 Wall S, Kunze ZM, Saboor S, et al. Identification of spheroplast-like agents isolated from tissues of patients with Crohn's disease and control tissues by polymerase chain reaction. J Clin Microbiol 1993; 31: 1241–45.
- 54 Autschbach F, Eisold S, Hinz U, et al. High prevalence of Mycobacterium avium subspecies paratuberculosis IS 900 DNA in gut tissues from individuals with Crohn's disease. Gut 2005; 54: 944–49.
- 55 Bernstein CN, Nayar G, Hamel A, Blanchard JF. Study of animal-borne infections in the mucosas of patients with inflammatory bowel disease and population-based controls. J Clin Microbiol 2003; 41: 4986–90.
- 56 Bernstein CN, Blanchard JF, Rawsthorne P, Collins MT. Population-based case control study of seroprevalence of Mycobacterium paratuberculosis in patients with Crohn's disease and ulcerative colitis. J Clin Microbiol 2004; 42: 1129–35.
- 57 Bull TJ, McMinn EJ, Sidi-Boumedine K, et al. Detection and verification of Mycobacterium avium subsp. paratuberculosis in fresh ileocolonic mucosal biopsy specimens from individuals with and without Crohn's disease. J Clin Microbiol 2003; 41: 2915–23.
- 58 Cho SN, Brennan PJ, Yoshimura HH, Korelitz BI, Graham DY. Mycobacterial aetiology of Crohn's disease: serologic study using common mycobacterial antigens and a species-specific glycolipid antigen from Mycobacterium paratuberculosis. Gut 1986; 27: 1353–56.
- 59 Clarkston WK, Presti ME, Petersen PF, et al. Role of Mycobacterium paratuberculosis in Crohn's disease: a prospective, controlled study using polymerase chain reaction. Dis Colon Rectum 1998; 41: 195–99.
- 60 Collins MT, Lisby G, Moser C, et al. Results of multiple diagnostic tests for Mycobacterium avium subsp. paratuberculosis in patients with inflammatory bowel disease and in controls. J Clin Microbiol 2000; 38: 4373–81.
- 61 Elsaghier A, Prantera C, Moreno C, Ivanyi J. Antibodies to Mycobacterium paratuberculosis-specific protein antigens in Crohn's disease. Clin Exp Immunol 1992; 90: 503–08.
- 62 Erasmus DL, Victor TC, van Eeden PJ, Falck V, van Helden P. Mycobacterium paratuberculosis and Crohn's disease. Gut 1995; 36: 942.
- 63 Fidler HM, Thurrell W, Johnson NM, Rook GA, McFadden JJ. Specific detection of Mycobacterium paratuberculosis DNA associated with granulomatous tissue in Crohn's disease. Gut 1994; 35: 506–10.

- 64 Lisby G, Andersen J, Engbaek K, Binder V. Mycobacterium paratuberculosis in intestinal tissue from patients with Crohn's disease demonstrated by a nested primer polymerase chain reaction. Scand J Gastroenterol 1994; 29: 923–29.
- 65 Murray A, Oliaro J, Schlup MM, Chadwick VS. Mycobacterium paratuberculosis and inflammatory bowel disease: frequency distribution in serial colonoscopic biopsies using the polymerase chain reaction. Microbios 1995; 83: 217–28.
- 66 Naser SA, Ghobrial G, Romero C, Valentine JF. Culture of Mycobacterium avium subspecies paratuberculosis from the blood of patients with Crohn's disease. Lancet 2004; 364: 1039–44.
- 67 Romero C, Hamdi A, Valentine JF, Naser SA. Evaluation of surgical tissue from patients with Crohn's disease for the presence of Mycobacterium avium subspecies paratuberculosis DNA by in situ hybridization and nested polymerase chain reaction. Inflamm Bowel Dis 2005; 11: 116–25.
- 68 Ryan P, Bennett MW, Aarons S, et al. PCR detection of Mycobacterium paratuberculosis in Crohn's disease granulomas isolated by laser capture microdissection. Gut 2002; 51: 665–70.
- 69 Suenaga K, Yokoyama Y, Okazaki K, Yamamoto Y. Mycobacteria in the intestine of Japanese patients with inflammatory bowel disease. Am J Gastroenterol 1995; 90: 76–80.
- 70 Suenaga K, Yokoyama Y, Nishimori I, et al. Serum antibodies to Mycobacterium paratuberculosis in patients with Crohn's disease. Dig Dis Sci 1999; 44: 1202–07.
- 71 Tiveljung A, Soderholm JD, Olaison G, Jonasson J, Monstein HJ. Presence of eubacteria in biopsies from Crohn's disease inflammatory lesions as determined by 16S rRNA gene-based PCR. J Med Microbiol 1999; 48: 263–68.
- 72 Walmsley RS, Ibbotson JP, Chahal H, Allan RN. Antibodies against Mycobacterium paratuberculosis in Crohn's disease. QJM 1996; 89: 217–21.
- 73 Hulten K, El Zimaity HM, Karttunen TJ, et al. Detection of Mycobacterium avium subspecies paratuberculosis in Crohn's diseased tissues by in situ hybridization. Am J Gastroenterol 2001; 96: 1529–35.
- 74 Nakase H, Nishio A, Tamaki H, et al. Specific antibodies against recombinant protein of insertion element 900 of Mycobacterium avium subspecies paratuberculosis in Japanese patients with Crohn's disease. Inflamm Bowel Dis 2006; 12: 62–69.

- 75 Schlesselman J. Case-control studies. Design, conduct, analysis. New York: Oxford University Press, 1982.
- 76 Tasara T, Stephan R. Development of an F57 sequence-based realtime PCR assay for detection of Mycobacterium avium subsp paratuberculosis in milk. Appl Environ Microbiol 2005; 71: 5957–68.
- 77 Cousins DV, Whittington R, Marsh I, Masters A, Evans RJ, Kluver P. Mycobacteria distinct from Mycobacterium avium subsp. paratuberculosis isolated from the faeces of ruminants possess IS 900-like sequences detectable IS 900 polymerase chain reaction: implications for diagnosis. Mol Cell Probes 1999; 13: 431–42.
- 78 van Schaik G, Schukken YH, Crainiceanu C, Muskens J, Van Leeuwen JA. Prevalence estimates for paratuberculosis adjusted for test variability using Bayesian analysis. Prev Vet Med 2003; 60: 281–95.
- 79 Traunmüller F. Etiology of Crohn's disease: do certain food additives cause intestinal inflammation by molecular mimicry of mycobacterial lipids? Med Hypotheses 2005; 65: 859–64.
- 80 Marks DJ, Harbord MW, MacAllister R, et al. Defective acute inflammation in Crohn's disease: a clinical investigation. *Lancet* 2006; 367: 668–78.
- 81 Greenstein RJ, Collins MT. Emerging pathogens: is Mycobacterium avium subspecies paratuberculosis zoonotic? Lancet 2004; **364**: 396–97
- 82 Donaghy JA, Totton NL, Rowe MT. Persistence of Mycobacterium paratuberculosis during manufacture and ripening of cheddar cheese. Appl Environ Microbiol 2004; 70: 4899–05.
- 83 Grant IR, Hitchings EI, McCartney A, Ferguson F, Rowe MT. Effect of commercial-scale high-temperature, short-time pasteurization on the viability of Mycobacterium paratuberculosis in naturally infected cows' milk. Appl Environ Microbiol 2002; 68: 602–07.
- 84 Grant IR, Ball HJ, Rowe MT. Incidence of Mycobacterium paratuberculosis in bulk raw and commercially pasteurized cows' milk from approved dairy processing establishments in the United Kingdom. Appl Environ Microbiol 2002; 68: 2428–35.